doi:10.18081/2333-5106/016-6/233-242

Effect of prepubertal and postpubertal long-term exposure to the mild dose of soybean seeds on the tissue and function of thyroid gland in albino rat

Siham M. Abdul-Ridha<sup>1\*</sup>, Abed H. Baraaj<sup>2</sup>, Salim R. Hamudi<sup>3</sup>

#### **Abstract**

The aim of this study is to investigate the effect of long-term exposure to the mild dose of soybean seeds on the tissue and some physiological parameters of the thyroid gland in the prepubertal and postpubertal life stages. Twenty four Sprague-Dawley albino male rats were divided into four groups (n=6); control rats at the prepubertal life stage; prepubertal rats treated with 20% soybean seeds of daily diet for 40 days after the weaning; normal control rats, at the postpubertal stage; and postpubertal rats, were treated with 20% soybean seeds. Morphmetrical, histological and physiological changes were examined. Consumption of mild dose of soybean seeds along the prepubertal life stage showed significant decrease (P<0.05) in the height of follicular cell, significant increase (P<0.05) in the diameter of follicular lumen and ratio of cold follicles, slightly non-significant decline in (T3 and T4) hormones levels and significant increment (P<0.05) in body weight, while at postpuberty, long-term exposure for the same dose of soybean seeds showed significant increase in the height of follicular cell (P<0.05), significant decrease (P<0.05) in the diameter of follicular lumen and ratio of cold follicles caused hyperactivity of the thyroid, significant decline (P<0.05) in (T3 and T4) hormones levels and slightly non-significant increment (P<0.05) in body weight. We concluded that the long-term exposure to the mild dose of the soybean affect adversely the tissue and function of the thyroid at both life stages, pre- and postpuberty.

**Keywords:** Soybean seeds; Tissue; Function; Thyroid; Puberty

\*Corresponding Author: Siham M. Abdul-Ridha. Electronic address: sihamm.alrehemi@uokufa.edu.iq

<sup>1</sup>Department of Anatomy and Histology, College of Medicine, Kufa University

<sup>2</sup>Department of Biology, College of Science, University of Baghdad

<sup>3</sup>Department of Pathology, College of Medicine

Received January 02, 2016; accepted May 02, 2016; published June 25, 2016

Copyright © 2016 SA. This is article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and

reproduction in any medium, provided the original work is properly cited. (cc) BY



#### Introduction

There is growing interest in the potential health threats posed by natural and anthropogenic substances in hum-an's and rodent's food, water and the environment which called endocrine-disrupting chemicals to the endocrine system [1].

From these diet contents and phytoestrogens, substances: which interfere with the absorption of synthetic thyroid hormone [2]. Soybean [Glycine max (L.) Merrill] is the most important dietary source of isoflavones, an important class of phytoestrogen [1]. The popularity and healthy importance of the soybean, which is the miracle crop of 20<sup>th</sup> century [3, 4] and the traditional staple of Asian diet, was promoted by positive findings between soy isoflavone consumption and lower incidences of diseases [5], due to the its many estrogenic, antioxidative, antiosteoporotic, anti-carcinogenic [6], antidiabetic activities, both in an animal model [7] and in human epidemiological study [8].

It is consumed just alone or in combination with other food products by human beings and as animal feed [9]. It represents an excellent source of high quality protein [10], which comprises 35-40% dry weight of the soybean [11, 12] and contains all amino acids essential to human nutrition, which make it unique among plant-based proteins [12].

Soybean contains on average 15-20% oil by weight [13] which characterized by its high content of polyunsaturated fats, less saturated fat and no cholesterol. Also, it contains

macronutrients such as carbohydrates [10] and a wide range of micronutrients and phytochemicals including phytic acid 1.0–2.2%, sterols 0.23–0.46%, and saponins 0.17–6.16% dry weight of the soybean [14], and the rest of the soybean components is vitamins, minerals [10], moister, ash and no sodium. So, it can be said that soybean contain all the nutrients needed in food [15].

Concerns have been expressed that soy iso-flavone intake adversely affects several organs, one of these organs is the thyroid [2]. The thyroid gland is one of the largest endocrine glands, locates in the neck bellow the larynx. It composed of two lobes, joined by an isthmus, each lobe consists of large numbers of closed spherical follicles [16]. The size of the thyroid follicles varies from species to species, for example, it is larger in man than in rat, it is generally quite small in newborn and increases progressively with age. The height of the follicular epithelium depends on its activity and is often inversely proportional to the diameter of follicular lumen. In a resting (cold) [17] inactive gland, the follicle becomes large, the surrounding cells become flattened with abundant and dense colloid [17 and 18], while in a hot (active) gland, under thyroid stimulating hormone(TSH) stimulation epithelial height increases and becomes columnar with a few, faint colloid and small size follicle [17].

The first description of goitrogenic effects of soybean was by McCarisson [19] who indicated that several investigators have reported induction of goiter in iodine-deficiency rats maintained on soybean diet. Goiter and hypothyroidism were reported in infants

fed with adapted soy formula without adequate iodine supply [20].

It was indicated that both genistein, and to a lesser extent daidzein (the most healthy important forms of isoflavones in soybean) were shown to strongly inhibit the activity of thyroid peroxidase (TPO) [21 and 22]. Also, It was indicated that isoflavones lead to immune dysfunction by causing potent stimulation of T and B cells mediated immunity due to induced structural changes in thyroid peroxidase [23], this result agreed with findings of Xiao, et al. [24] which revealed that isoflavones suppress the binding ability of hepatic thyroid hormone receptor to the thyroid hormone response element of the target genes. Similarly, it was revealed that both genistein and daidzein impaired thyroglobulin thyroid (Tg)and hormones synthesis of Orx middle-aged rats by decrease the expression of Tg and TPO genes [18].

## **Method and Material**

## Preparation of Soybean Pellet

Soybean seeds which were obtained as defatted seeds from the local fodder the Al-Sinak markets in Street/ Baghdad, were powdered by seed grinder, and this soybean powder used in the preparation of soybean pellet according to the study [25] as following: 50% bran, 25% white flour, 20% soybean powder and 5% sugar, vitamin, soybean oil and salt. These components were mixed, kneaded to make a cylinder blocks similar to the normal rodent pellet and dried under the sun temperature for a few hours. The resulted pellet represented the soybean pellet .

Animals care and experimental design Twenty four Sprague-Dawley (Rattus norvegicus) albino male rats were used in the present study as a mammalian model, purchased from the College of Medicine/Baghdad University and housed in the animal house of the College of Science for the same University, kept in plastic cages with a metal network cover under climate controlled conditions of the animal house with temperature 25±2°C and

12:12 light and dark cycle.

They were divided into four groups, each of six rats, the experimental groups of rats (G1, G2) were treated directly after their weaning (prepubertal stage) in a daily consu-mption average 12-15 g, weighing 33-40 g, while the experimental groups of rats (G3, G4) were treated after their puberty (postpubertal stage) in daily consumption average 18-20 g, weighing 160-190g. All the experimental animals pelleted for 40 days as following: groups (G1 and G3), normal control rats, were provided with water and food ad libitum, while the groups (G2 and G4) were treated with (20%) soybean seeds from the daily diet [26.]

# Collection of blood samples and dissection the animals

At the end of the experiment, the animals were weighed, fully anaesthetized by diethyl ether for several minutes and the blood samples were obtained by heart puncture. Sera were separated by centrifugation 3000 rpm for 15 min, and then they were kept in  $-20^{\circ}$ C until using them. The animals

ticle doi:10.18081/2333-5106/016-6/233-242

were dissected and the thyroid gland was excised, washed and fixed in neutral buffered 10% formalin for the histological preparation.

# Histopathological examination

The fixed tissues of the thyroid in the neutral buffered 10% formalin were washed with running tap water. dehydrated through graded alcohol series (50-100%), cleared with xylene, infiltrated and embedded in paraffin wax. The paraffin blocks were sectioned in thickness 4-5  $\mu$ , stained with Hematoxylin and Eosin stain (H&E) and Toluidine Blue stain (T.B.) and finally mounted by Canada Balsam [27], then the sections were examined histologically.

## Physiological Examination

It was represented by the enzyme immunoassay tests (ELISA) for the quantitative determination of concentrations of thyroid gland hormones (T3 and T4). Measurement the level of T3 concentration was performed according to the Triiodothyronine (T3) enzyme immunoassay test kit, catalog number: BC- 1005, from BioCheck, Inc, 323 Vintage Park Dr., Foster City, CA 94404, and measurement the level of T4 concentration was performed according to the total thyroxine (T4) enzyme immunoassay test kit, catalog number: BC- 1007, from BioCheck, Inc ,323 Vintage Park Dr., Foster City, CA 94404.

# Morphometrical examination

The resulted sections from the histological preparation were examined morphometrically to determination the height of follicular cell and the diameter of follicular lumen using Motic Image Plus version 2.0 software program [28], while the ratio of the cold follicles was calculated according to the study [29.[

## Statistical analysis

It was performed by using Statistical Package of Social Sciences (SPSS), version 2.0, (ITS, Los Angeles) computer software. Differences between groups were analyzed by using an analysis of variance (ANOVA), P<0.05 was regarded as statistically significant [30].

#### **Results and Discussion**

## Weight Measurement

The statistical analysis of the present study **Tab. 1**, indicated that the soybean consumption significant caused increment (P<0.05) in body weight in slightly non-significant (G2)and increment (P<0.05) in body weight in (G4) compared to the control groups due to the high soybean content of protein which comprises 35-40% from its dry weight, its amino acids are important to muscle synthesis and consequently increase the weight. This result agreed with the results of another previous study [31 and 32]. Also, the study [33] indicated that soybean proteins caused a reduction of the proteolysis myofibrillar protein in skeletal muscles; this may lead to increase the weight.



Parameters	Means ± SE			
	Prepuberty		Postpuberty	
	Control (G1)	20% soybean <b>(G2)</b>	Control (G3)	20% soybean <b>(G4)</b>
Body weight (g)	$A$ 122.67 $\pm$ 4.62	$\mathbf{B}$ 174.17 ± 3.24	A 269 ± 10.67	$A \\ 309.33 \pm 6.53$
T3 level (ng/ml)	A 3.836 ± 0.26	A 3.298 ± 0.45	$A$ 3.608 $\pm$ 0.27	$\mathbf{B}$ 2.534 ± 0.15
T4 level (μg/dl)	$A \\ 11.33 \pm 0.31$	A 9.80 ± 1.73	A 11.79 ± 0.25	$\mathbf{B}$ 9.46 ± 0.58
Height of follicular cell (µm)	<b>A</b> 123.28 ±4.75	<b>B</b> 97.61 ± 5.03	$A$ 129.62 $\pm$ 3.86	$\frac{\mathbf{B}}{170.32 \pm 7.58}$
Diameter of follicular lumen (μm)	A 322.93±31.07	$\mathbf{B} \\ 460.09 \pm 40.49$	$A 500.98 \pm 32.86$	<b>B</b> 376.36 ±33.39
Ratio of cold follicles	$A \\ 16.23 \pm 0.689$	$\mathbf{B}$ 25.10 ± 0.955	$A 21.40 \pm 1.89$	$\mathbf{B}$ $5.96 \pm 0.34$

SE= standard error

Different letters (A and B) are significant at (P < 0.05)

## Physiological Examination

Results of the statistical analysis of the present study **Tab. 1**, revealed a slightly non-significant decline P<0.05 in T3 and T4 levels in the (G2) and significant decline P<0.05 in T3 and T4 levels in the (G4) compared to the control groups. This reduction in thyroid hormones levels may be resulted due to the inhibition of thyroper-oxidase- (TPO) catalyzed reactions which essential to thyroid hormone synthesis by the soybean compounds and consequently decreases thyroid hormones levels. In the presence of iodide ion, genistein and daidzein blocked TPO-catalyzed tyrosine iodination by acting as alternate substrates, yielding mono, di, and triiodoisoflavones [34].

Similarly, the study [18] showed decrease in expression of thyroglobulin

(Tg) and thyroperoxidase (TPO) genes at level (P<0.05) due to soybean consumption. On the other hand, it was indicated that soybean inhibited iodide absorption and consequently affected thyroid hormone synthesis [2]. The results of the present study agreed with the results of another previous studies [35, 36, 18].

Coordinating with the morphmetrical and histopathological findings, the treated rats with soybean which revealed the reduction in thyroid hormones T3 and T4 showed either increase in the cold (inactive) follicles ratio or hypertrophy of the follicular epithelium **Tab. 1**, **Fig. 1b**, **d** and **Fig. 2b**, **d**, both cases indicted that the thyroid gland was impaired and consequently may had depressed activity of this gland to produce the thyroid hormones.

doi:10.18081/2333-5106/016-6/233-242

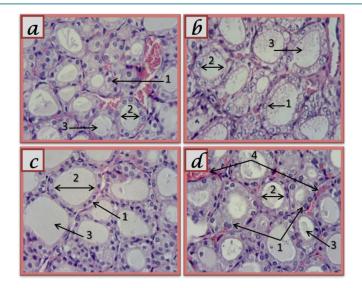


Figure 1.

Sections of thyroid gland from the experimental groups of rats (a): control at prepuberty, in which showing normal follicles structure lined by cuboidal epithelium with normal amount of colloid material, (b): treated with 20% soybean at prepuberty, in which showing follicles lined by cuboidal to flat epithelial cells, (c): control at postpuberty, in which showing normal follicles structure lined by cuboidal epithelium with normal amount of colloid material and (d): treated rats with 20% soybean at postpuberty, in which showing increase in the height of epithelium, decrease in the follicular lumen, less colloid material, congesting of the blood vessels (H&E) 400x

1- Follicular epithelium 2- Follicular lumen 3- Colloid material4- blood vessels

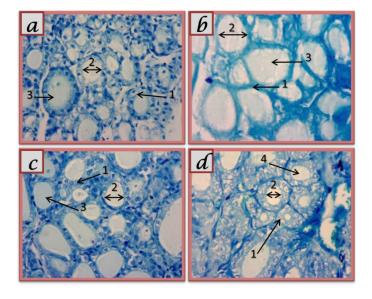


Figure 2.

Sections of thyroid gland from the experimental groups of rats (a): control at prepuberty, in which showing normal follicles structure lined by cuboidal epithelium with normal amount of colloid material, (b): treated with 20% soybean at prepuberty, in which showing follicles lined by flat epithelial cells, (c): control at postpuberty, in which showing normal follicles structure lined by cuboidal epithelium with normal amount of colloid material and (d): treated rats with 20% soybean at postpuberty, in which showing increase in the height of epithelium, decrease in the follicular lumen, consumed colloid material (T.B.) 400x.

1- Follicular epithelium 2- Follicular lumen 3- Colloid material 4- Consumed colloid



## Histopathological examination

The statistical analysis of the present study morphometry Tab. 1, revealed significant decrease (P<0.05) in the height of follicular cell, significant increase (P<0.05) in the diameter of follicular lumen and significant increase (P<0.05) in the ratio of cold follicles in the treated group of rats at prepuberty (G2) compared to the control group, while the treated group at postpuberty showed significant increase (G4) (P<0.05) in the height of follicular cell, significant decrease (P<0.05) in the diameter of follicular lumen and significant decrease (P<0.05) in the ratio of cold follicles compared to the control group.

Coordinating with the morphometry Tab. 1, histologically Fig. 1b, 2b, longterm exposure to the mild dose of soybean at prepuberty (G2) showed an inhibitor effect, which represented by several histological findings such as flattening of epithelial cells, increase in the diameter of follicular lumen, increase in the ratio of cold follicles and accumulation of colloid material, while at postpuberty Fig. 1d, 2d, and due to the feedback mechanism more secretion of TSH which led to hyperactivity, soybean consumption caused increase in the height of follicular cell "cuboidal to columnar epithelial cell", decrease in the diameter of follicular lumen and ratio of cold follicles, less amount of colloid material and congesting of the blood vessels.

Inhibitor effect of soybean seeds varied in the prepuberty from the postpuberty, at prepuberty, the endocrine gland (pituitary gland) is not well formed and has a minor effect [37] and not functional established, therefore, no more TSH secretion led to decrease the activity rate of the thyroid gland **Fig. 1b, 2b,** while at postpuberty, TSH secretion increased due to the feedback mechanism causing hyperactivity of the thyroid led to the hypertrophy of the follicles (figures 1d and 2d), this excessive TSH secretion stimulated abnormal thyroid growth and may had developed the goiter [37].

The study [34] revealed that the inhibition of thyroid hormone synthesis can induce goiter in rodents, which agreed with the result of the present study, as well, the present study result agreed with the result of the study [36] which revealed that the epithelial height and index of activation rate increased, while the luminal colloid and the thyroid hormones decreased after treating the Wistar male rats with 10 mg/kg of either genistein or daidzein for three weeks.

## In conclusions

From this study, it is concluded that the long-term exposure to the mild dose of soybean increased body weight, reduced thyroid hormones levels at postpuberty, reduced the activity of the thyroid and impaired its tissue at prepuberty and caused hyperactivity of the gland at postpuberty.

# **Competing interests**

Authors declare that We have no competing interests.

# Research Article doi:10.18081/2333-5106/016-6/233-242

## **Authors' contributions**

SA and AB were involved in design of the study, involved in data collection and drafting of the final manuscript. SH involved in conception and design of the study, and offered a critical review of the manuscript for intellectual content. All the authors have read and approved the final version of the manuscript.

#### References

- 1. Cederroth CR, Zimmermann C, Nef SS. Phytoestrogens and their impact on reproductive health. Mol Cell Endocrinol 22;355(2):192-200.
- 2. Messina M, Redmond G. Effects of soy protein and soybean isoflavones on thyroid function in healthy adults and hypothyroid patients. Official journal of the American thyroid association 2006; **16**(3):249-258.
- 3. Hymowitz T, Newell CA. Taxonomy, speciation, domestication, dissemination, germplasm resources and variation in the genus Glycine. p. 149-225. In: R.J. Summerfields and A.H. Bunting (ed.), Advances in legume science. Royal Botanical Gardens, Kew, London, 1978.
- 4. Govindarao CN. Characterization of soybean [Glycine max (L.) Merrill. varieties through morphological, chemical, molecular markers and image analyzer. M. Sc. Thesis. College of Agriculture, University of Agricultural Sciences, Dharwad, 2010.
- 5. Shu XO, Zheng Y, Cai H, et al. Soy food intake and breast cancer survival. JAMA 2009:302:2437-2443.
- 6. Retana-Marquez S, Hernandez H, Flores JA, et al. Effect of phytoestrogens on mammalian reproductive physiology. Tropical and Subtropical Agroecosystems 2012;**15**(1):129-145.

- 7. Dyrskog EU, Jeppesen PB, Colombo M, Abudula R, Hermansen K. Preventive effects of a soy-based diet supplemented with stevioside on the development of the metabolic syndrome and type 2 diabetes in Zucker diabetic fatty rats. Metabolism 2005;54:181-1188.
- 8. Villegas R, Gao YT, Yang G, et al. Legume and soy food intake and the incidence of type 2 diabetes in the Shanghai Women's Health Study. American Journal of Clinical Nutrition 2008;**87**:162-167.
- 9. Ekaluo UB, Udoh PB, Ikpeme EV. Effect of soybean (Glycine max L.) on the milieu of male International Journal of Recent Scientific Research 2012;3(8):722-724.
- 10. Asif M, Acharya M. Phytochemicals and nutritional health benefits of soy plant. International Journal of Nutrition, Pharmacology, Neurological Diseases 2013;3(1):64-69.
- 11. Torres N, Torre-Villalvazo I, Tovar AR. Regulation of lipid metabolism by soy protein and its implication in diseases mediated by lipid disorders. Journal of Nutritional Biochemistry 2006;17:365-373.
- 12. Singh P, Kumar R, Sabapathy SN, Bawa AS. Functional and edible uses of soy Comprehensive protein products. Reviews in Food Science and Food Safety 2008;7:14-28.
- 13. Sugano M. Nutritional implications of soybean. p. 3-16. In: M. Sugano (ed.), Soy in Health and Disease Prevention, CRC Press: Boca Raton, FL, 2006.
- 14. Kang J, Badger TM, Ronis MJ, Wu X. Non-isoflavone phytochemicals in soy and their health effects. Journal of Agricultural and Food Chemistry 2010;**58**:8119-8133.
- 15. Apriletti JW, Riberio RC, Wagner RL, et al. Molecular and structural biology of thyroid hormone receptors. Clin. Exp.

- Pharmacol. Physiol Suppl 1998;25:S2-11.
- Hamudi SR. Involution of thyroid hyperplasia. Ph.D. Thesis. Catholic University of Louvain, England, 1986.
- 17. Šošić-Jurjević B, Filipović B, Wirth EK, et al. Soy isoflavones interfere with thyroid hormone homeostasis in orchidectomized middle-aged rats. *Toxicology and Applied Pharmacology* 2014;**142**:1-11.
- 18. McCarisson R. The goitrogenic action of soybean and groundnut. *Indian Journal of Medical Research* 1993; **21**:179-181.
- 19. Van Wyk JJ, Arnold MB, Wynn J, Pepper F. The effects of a soybean product on thyroid function in humans. *Pediatrics* 1959;**24**:752-760.
- 20. Chang HC, Doerge DR. Dietary genistein inactivates rat thyroid peroxidase in vivo without an apparent hypothyroid effect. *Toxicology and Applied Pharmacology* 2000;**168**:244-252.
- 21. Doerge DR, Sheehan D. Goitrogenic and estrogenic activity of soy isoflavones. *Environmental Health Perspectives* 2002;**110**(3):349-353.
- 22. Chen A, Rogan WJ. Isoflavones in soy infant formula: a review of evidence for endo-crine and other activity in infants. *Annual Review of Nutrition* 2004;**24**:33-54.
- 23. Xiao CW, L'Abbé MR, Gilani S, Cooke G, Curran I, Papademetriou SA. Dietary soy protein isolate and isoflavones modulate he-patic thyroid hormone receptors in rats. *Journal of Nutrition* 2004;**134**:743-749.
- 24. Al-Hilfy JH. Effect of soybean seeds on liver function and apoptosis in liver, spleen and thymus in male rats treated with uranyl nitrate. Ph.D. Thesis. College of Science, University of Baghdad, Iraq, 2007.

- 25. Tovar AR, Murguia F, Cruz C, et al. A soy protein diet alters hepatic lipid metabolism, gene expression and reduces serum lipids and renal fibrogenic cytokins in rats with chronic nephritic syndrome. *J. Nutr* 2002; **132**(9):2562-9.
- 26. Suvarna SK, Layton C, Bancroft JD. Bancroft's theory and practice of histological techniques, 7th ed. Elsevier Lim, China, 2013.
- 27. Salih LA. The teratogen effects of pregabalin drug on the cerebellar development of albino rat embryos. Ph.D. Thesis, College of Science, University of Baghdad, Iraq.
- Al-Sudany NM. Thyroid hormones inhibitors and spermatogenesis. Ph.D. Thesis. College of Science, Al-Mustansiriyah University, Iraq, 2004.
- 29. Calstatela. IBM SPSS statistics 20/part4: Chi-square and ANOVA. ITS, Los Angeles, 2013.
- 30. Amer N. Effects of Soybean Seed on Glucose Levels, Lipid Profiles and Histological Structures of the Liver in Alloxan-Induced Diabetic Albino Rats. *Tikrit Journal of Pure Science* 2012; **17**(2):1-5.
- 31. Baraaj AH. Treatment of diabetic male rats with soybean [Glycine max (L.)] seeds. Histological and biochemical study. *Journal of the collage of basic education* 2014;**83**(20):655-670.
- 32. Tada O, Yokogoshi H. Effect of different dietary protein composition on skeletal muscle atrophy by suspention hypokinesia/ hypodynamia in rats. *Journal of Nutritional Science and Vitaminology* 2002;**48**(2):115-119.
- 33. Divi RL, Chang HC, Doerge DR. Antithyroid isoflavones from soybean: isolation, characterization, and mechanisms of action. *Biochemical Pharmacology* 1997;54(10):1087-1096.
- 34. Surks MI, Hollowell JG. Age-specific distribution of serum thyrotropin and antithyroid antibodies in the US

doi:10.18081/2333-5106/016-6/233-242

- population: implications for the prevalence of subclinical hypothyroid-dism. *Journal of Clinical Endocrinology and Metabolism* 2007; **92**:4575-4582.
- 35. Šošić-Jurjević B, Filipović B, Ajdžanović V, et al. Suppressive effects of genistein and daidzein on pituitary-thyroid axis in Orchidectomized middle-aged rats. *Journal of Experimental Biology and Medicine* 2010;**235**(5):590-598.
- 36. Sharpe RM, Fraser HM, Brougham MF, et al. Role of the neonatal period of pituitary-testicular activity in germ cell proliferation and differentiation in the primate testis. *Human Reproduction* 2003;**18**(10):2110-2117.
- 37. Vanderpump MP. The epidemiology of thyroid disease. *British Medical Bulletin* 2011;**99**:39-51.

